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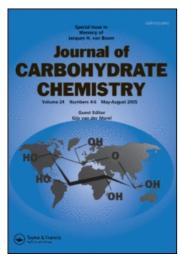
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SYNTHETIC STUDIES ON SIALOGLYCOCONJUGATES 91: TOTAL SYNTHESIS OF GANGLIOSIDES GD1c AND GT1a

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ABSTRACT

A first total synthesis of gangliosides GD1c and GT1a containing Neu5Aca(2→8) Neu5Aca(2→3)Gal residue in their non-reducing terminal is described. Condensation of methyl O-[methyl 5-acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-Dglycero-a-D-galacto-2-nonulopyranosylono-1',9-lactone) -4,7- di-O-acetyl-3,5 -dideoxy-Dglycero-α-D-galacto-2-nonulopyranosylonate]-(2 → 3)-2,4,6-tri-O-benzoyl-1-thio-β-D-galactopyranoside (1) with 2-(trimethylsilyl)ethyl O-(2-acetamido-4,6-O-benzylidene-2-deoxy- β -D-galactopyranosyl) - (1 \rightarrow 4) -O -(2,3,6-tri-O-benzyl-β-D-galactopyranosyl)-(1 \rightarrow 4) -2,3,6tri-O-benzyl-β-D-glucopyranoside (2) or 2-(trimethylsilyl)ethyl O-(2-acetamido-6-Obenzyl-2-deoxy-β-D-galactopyranosyl)-(1→4)-O-[methyl 5-acetamido-4,7,8,9-tetra-Oacetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2 \rightarrow 3)$]-O-(2,6-di-Obenzyl- β -D-galactopyranosyl) - (1 \rightarrow 4) - 2,3,6-tri-O-benzyl- β -D-glucopyranoside (3) in the presence of dimethyl(methylthio)sulfonium triflate (DMTST) gave the corresponding hexaand heptasaccharide derivatives 4 and 5, respectively. These oligosaccharides were converted into the a-trichloroacetimidates 10 and 11 via reductive removal of the benzyl groups and/or benzylidene group, O-acetylation, selective removal of (trimethylsilyl)ethyl group and treatment with trichloroacetonitrile, which, on coupling with 2-azidosphingosine derivatives 12 or 13, gave the β-glycosides 14 and 15, respectively. Finally, 14 and 15 were transformed, via selective reduction of the azido group, coupling with octadecanoic acid and removal of all protecting groups, into the title gangliosides GD1c 18 and GT1a 19.

INTRODUCTION

Ganglio-series gangliosides are distinguished from other gangliosides in that they contain gangliotriosylceramide $[GalNAc\beta(1\rightarrow 4)Gal\beta(1\rightarrow 4)Glc\beta(1\rightarrow 1)Cer]$ or gangliotetraosylceramide [Gal $\beta(1\rightarrow 3)$ GalNAc $\beta(1\rightarrow 4)$ Gal $\beta(1\rightarrow 4)$ Glc $\beta(1\rightarrow 1)$ Cer] as the neutral sugar chain backbones in their molecules, and sialic acids link at C-3 of Gal, C-6 of GalNAc and/or C-8 of Neu5Ac. These gangliosides are found in the central nervous system, 2-5 and are considered to play various important roles in biological systems. 6-12 In order to elucidate the functions of gangliosides in detail the pure compounds are required, because biologically derived gangliosides are polymorphous at the ceramide unit and available only in very limited quantity. Therefore, it is important to develop a facile chemical synthesis of these complex glycolipids. We have reported systematic syntheses 13-23 of various types of gangliosides and their analogs in order to elucidate structure-function relationships at the molecular level. We describe herein the synthesis of gangliosides GD1c and GT1a which contain the Neu5Aca(2→8)Neu5Aca(2→3)Gal residue in their non-reducing terminal. Gangliosides GD1c and GT1a were first isolated from murine thymoma by Bartoszewicz et al.24 and from human brain by Ando et al.,25 respectively. The biological functions of these gangliosides have not been reported yet because of the reason described above.

RESULTS AND DISCUSSION

For the synthesis of the desired gangliosides GD1c and GT1a we have employed methyl O-[methyl 5-acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonuropyranosylono-1,9'-lactone)-4,7-di-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate]-(2 \rightarrow 3)-2,4,6-tri-O-benzoyl-1-thio- β -D-galactopyranoside²⁰ (1) as a key glycosyl donor, and 2-(trimethylsilyl)ethyl O-(2-acetamido-4,6-O-benzylidene-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2,3,6-tri-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside²⁶ (2) and 2-(trimethylsilyl)ethyl O-(2-acetamido-6-O-benzyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-[methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonuropyrano-

AcO	ÇOOMe OBz	XO-	OR3 OR3	0R ³ R ³ O R ²
AcOIII Te O	BzO OBz	O C ORS	5	

	R ¹	R ²	R^3	R ⁴	R^5	X	
4	OSE	Н	Bn	benzy	lidene	Bn	
5	OSE	H	Bn	Н	Bn	SA	
6	OSE	Н	Ac	Ac	Ac	Ac	
7	OSE	Н	Ac	Ac	Ac	SA	
8	Н, ОН		Ac	Ac	Ac	Ac	
9	н, он		Ac	Ac	Ac	SA	
10	H OC(=NH)CCI ₃		Ac	Ac	Ac	Ac	
11	H OC(=NH)CCI ₃		Ac	Ac	Ac	SA	

SE = 2-(trimethylsilyl)ethyl Bn = benzyl Bz = benzoyl

HO
$$C_{13}H_{27}$$
OR

12 R = Bz
13 R = TBDPS

18 ganglioside GD1c

19 ganglioside GT1a

TBDPS = tert-butyldiphenylsilyl

sylonate)- $(2\rightarrow 3)$]-O-(2,6-di-O-benzyl- β -D-galactopyranosyl)- $(1\rightarrow 4)$ -2,3,6-tri-O-benzyl- β -D-glucopyranoside (3) as the well designed, glycosyl acceptors, suitable for the preparation of the hexa- and heptasaccharide derivatives 4 and 5.

By further processing, according to our usual procedure, ¹³ the resulting oligosaccharide intermediates 4 and 5 could be transformed into the end products by introduction of a ceramide moiety.

Glycosylation of **2** or **3** with **1** in the presence of dimethyl(methylthio)sulfonium triflate 13,23 (DMTST) and molecular sieves 4\AA (MS- 4\AA) gave the desired hexasaccharide **4** (56%) and heptasaccharide **5** (31%), respectively. Reductive removal (10% Pd-C) of the benzyl and/or benzylidene groups of **4** and **5** in ethanol-acetic acid, and subsequent *O*-acetylation gave the corresponding per-*O*-acyl-oligosaccharides **6** and **7** in good yields, respectively. The observed chemical shift and coupling constants for H-4c of the GalNAc residue (δ 5.53, J₃,4 = 3.5 Hz for **6**; δ 5.50, J₃,4 = 3.4 Hz for **7**) in **6** and **7** indicated the glycosylated position to be the 3-OH. Treatment 27 of **6** and **7** with trifluoroacetic acid in dichloromethane for 30 min at room temperature gave the 1-hydroxy compounds **8** (75%) and **9** (83%). When treated 28 with trichloroacetonitrile in dichloromethane in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) for 1 h at 0 °C, compounds **8** and **9** gave the corresponding α -trichloroacetimidates **10** (96%) and **11** (89%); significant signals of the Glc residue in the 1 H NMR spectra were a one-proton doublet (δ 6.43, J₁,2 = 3.7 Hz, for **10**; δ 6.48, J₁,2 = 3.7 Hz, for **11**), indicating the anomeric configuration of the imidates to be α .

Glycosylation of (2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol²⁹ (12) with 10, in dichloromethane in the presence of trimethylsilyl trifluoromethanesulfonate (TMSOTf) and molecular sieves 4Å (AW-300 type) for 10 h at 0 °C gave the desired β -glycoside 14 in 53% yield. Selective reduction^{29b,30} of azido group in 14 with hydrogen sulfide in aq 83% pyridine and subsequent condensation with octadecanoic acid, using 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (WSC) in dichloromethane, for 12 h at room temperature gave the protected ganglioside GD1c 16 in 82% yield. Finally, O-deacylation of 16 with sodium methoxide in methanol and subsequent saponification of the methyl ester and lactone group, yielded ganglioside GD1c (18) in almost quantitative yield.

On the other hand, condensation of 11 with (2S,3R,4E)-2-azido-3-O-(tert-butyldiphenylsilyl)-4-octadecene-1,3-diol³¹ (13) in dichloromethane, in the presence of TMSOTf, gave the desired β-glycoside 15 in 65% yield. Selective reduction³² of the azido group in 15 with triphenylphosphine in benzene-water for 24 h at 30 °C gave the amine, which was treated with octadecanoic acid in the presence of WSC in dichloromethane to give the protected ganglioside GT1a 17 in 95% yield. Finally, removal³³ of the *tert*-butyldiphenylsilyl group in 17 with M tetrabutylammmonium fluoride, O-deacylation, and saponification of the methyl ester and lactone group, yielded the desired ganglioside GT1a (19) in 75% yield after chromatography on a column of Sephadex LH-20. The ¹H NMR data of the products thus obtained are consistent with the structures assigned.

In conclusion, an efficient total synthesis of gangliosides GD1c and GT1a containing the Neu5Ac $\alpha(2\rightarrow 8)$ Neu5Ac $\alpha(2\rightarrow 3)$ -Gal residue at their non-reducing terminal was achieved by use of the key glycosyl donor 1, indicating its usefulness for the syntheses of polysialoglycoconjugates.

EXPERIMENTAL

General procedures. Optical rotations were determined with a Union PM-201 polarimeter at 25 °C, and IR spectra were recorded with a Jasco IRA-100 spectrophotometer. ¹H NMR spectra were recorded at 270 MHz with a Jeol JNM-GX 270 spectrometer and at 200 MHz with a Varian VXR spectrometer, and the NMR data were confirmed by use of decoupling techniques. Preparative chromatography was performed on silica gel (Fuji Silysia Co. 300 mesh) with the solvent systems specified. Concentrations were conducted in vacuo.

2-(Trimethylsilyl)ethyl O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyra-nosylono-1',9-lactone)-4,7-di-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate] - (2 \rightarrow 3)-O-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2-acetamido-4,6-O-benzylidene-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2,3,6-tri-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-

tri-O-benzyl-β-D-glucopyranoside (4). To a solution of methyl 5-acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylono-1',9-lactone)-4,7-di-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate]- $(2\rightarrow 3)$ -2,4,6-tri-O-benzoyl-1-thio- β -D-galactopyranoside²⁰ (1; 500 mg, 0.37) and 2-(trimethylsilyl)ethyl O-(2-acetamido-4,6-O-benzylidene-2-deoxy-β-Dgalactopyranosyl)- $(1 \rightarrow 4)$ -O-(2,3,6-tri-O-benzyl- β -D-galactopyranosyl)- $(1 \rightarrow 4)$ -2,3,6-tri-O-benzyl-β-D-glucopyranoside (2; 300 mg, 0.22 mmol) in CH₂Cl₂ (5 mL) were added molecular sieves 4Å (1.2 g) and the mixture was stirred for 5 h at room temperature and cooled to 0 °C. Dimethyl(methylthio)sulfonium triflate (DMTST; 370 mg, 0.74 mmol) was added, with stirring, to the mixture, and the stirring was continued for 2 days. After reaction was over, the precipitates were filtered off and washed with CH2Cl2. The filtrate and washings were combined, and the solution was successively washed with M Na2CO3 and water, dried (Na2SO4) and concentrated. Column chromatography (40:1 CH2Cl2-MeOH) of the residue on silica gel (30 g) gave 4 (330 mg, 56%) as an amorphous mass: $[\alpha]_D + 10.9^\circ$ (c 1.3, CHCl₃); IR (KBr) 3400-3100 (OH and NH), 1730 and 1230 (ester), 1650 and 1540 (amide), 860 and 840 (Me₃Si), 710 and 700 cm⁻¹ (phenyl); ¹H NMR (CDCl₃) δ 1.00 (m, 2 H, Me₃SiCH₂CH₂), 1.60 (t, 1H, J_{gem} = 13.0 Hz, H-3fax), 1.86-2.11 (9s, 27H, 6AcO, 3AcN), 2.45 (m, 1H, H-3eeq), 3.19 (s, 3H, MeO), 4.98 (t, 1H, $J_{1,2} = J_{2,3} = 7.9 \text{ Hz}, \text{ H-2d}, 5.12 \text{ (m, 1H, H-4e)}, 5.18 \text{ (d, 1H, } J_{7,8} = 8.5 \text{ Hz}, \text{ H-7e)},$ 5.35 (dd, 1H, $J_{6,7} = 1.4$ Hz, $J_{7,8} = 9.1$ Hz, H-7f), 5.56 (m, 1H, H-4f), 5.72 (d, 1H, $J_{3.4} = 3.5 Hz, H-4d), 7.12-8.12 (m, 50H, 10Ph).$

Anal. Calcd for C₁₃₆H₁₅₅N₃O₄₅Si (2579.8): C, 63.32; H, 6.06; N, 1.63. Found: C, 63.25; H, 5.86; N, 1.51.

2-(Trimethylsilyl)ethyl O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyra-nosylono-1',9-lactone)-4,7-di-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate] - (2 \rightarrow 3)-O-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-(2-acetamido-6-O-benzyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-[methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-O-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-galactopyranoside (5).

Glycosylation of 2-(trimethylsilyl)ethyl O-(2-acetamido-6-O-benzyl-2-deoxy-β-D-galactopyranosyl)-(1 \rightarrow 4)-O-[methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonuropyranosylonate)-(2 \rightarrow 3)]-O-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (3; 175 mg, 0.12 mmol) with 1 (250 mg, 0.18 mmol), as described for 4, gave 5 (103 mg, 31%) as an amorphous mass: [α]D +4.0° (c 1.0, CHCl3); IR (KBr) 3400-3100 (OH and NH), 1730 and 1230 (ester), 1650 and 1540 (amide), 860 and 840 (Me3Si), 710 and 700 cm⁻¹ (phenyl); ¹H NMR (CDCl3) δ 1.00 (m, 2H, Me3SiCH₂CH₂), 1.83-2.17 (14s, 42H, 10AcO, 4AcN), 2.36-2.60 (m, 3H, H-3e-geq), 3.60 (s, 3H, MeO), 7.18-8.10 (m, 45H, 9Ph).

Anal. Calcd for C₁₄₉H₁₇₈N₄O₅₈Si (2965.1): C, 60.36; H, 6.05; N, 1.89. Found: C, 60.18; H, 5.79; N, 1.85.

2-(Trimethylsilyl)ethyl O-[Methyl 5-Acetamido-8-0-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylono-1',9-lactone)-4,7-di-O-acetyl-3,5-dideoxy-D-glycero-\alpha-D-galacto -2-nonulopyranosylonate] - (2→3) - O- (2,4,6-tri-O-benzoyl-β-D-galactopyranosyl) - $(1 \rightarrow 3)$ - O- (2-acetamido-4, 6 - di-O-acetyl-2- deoxy- β -D-galactopyranosyl) - $(1 \rightarrow 4)$ - O- (2,3,6 - tri - O-acetyl - β - D - galactopyranosyl) - $(1 \rightarrow 4)$ - 2,3,6tri-O-acetyl-β-D-glucopyranoside (6). A solution of 4 (300 mg, 0.11 mmol) in AcOH (5 mL) and EtOH (5 mL) was hydrogenated in the presence of 10% Pd-C (300 mg) for 48 h at 45 °C, the catalyst removed by filtration and the solution concentrated. The residue was acetylated with Ac₂O (1 mL) and pyridine (2 mL) for 10 h at 45 °C. The mixture was concentrated, and a solution of the residue in CH2Cl2 was successively washed with 2 M HCl and M Na₂CO₃, dried (Na₂SO₄) and concentrated. Column chromatography (25:1 CH2Cl2-MeOH) of the residue on the silica gel gave 6 (272 mg, quantitative) as an amorphous mass: [a]p -6.2° (c 1.0, CHCl3); IR (KBr) 3200 (NH), 1730 and 1230 (ester), 1650 and 1540 (amide), 860 and 840 (Me₃Si), and 710 cm⁻¹ (phenyl); ${}^{1}H$ NMR (CDCl₃) δ 1.03 (m, 1H, Me₃SiCH₂CH₂), 1.63 (t, 1H, J_{gem} = 12.5 Hz, H-3fax), 1.88-2.22 (17s, 51H, 14AcO, 3AcN), 2.52 (m, 1H, H-3eeq), 3.46 (s, 3H, MeO), 5.04 (d, 1H, $J_{7.8} = 8.2$ Hz, H_{7e}), 5.07 (m, 1H, H_{4e}), 5.34 (dd, 1H, $J_{6.7} = 2.0$ Hz, $J_{7,8} = 8.9$ Hz, H-7f), 5.53 (d, 1H, $J_{3,4} = 3.5$ Hz, H-4c), 5.70 (d, 1H, $J_{3,4} = 3.6$ Hz, H-4d), 7.23-8.10 (m, 15H, 3Ph).

Anal. Calcd for C₁₀₃H₁₃₁N₃O₅₃Si (2287.2): C, 54.09; H, 5.77; N, 1.84. Found: C, 53.85; H, 5.73; N, 1.70.

2-(Trimethylsilyl)ethyl O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-a-D-galacto-2-nonulopyranosylono-1',9-lactone)-4,7-di-O-acetyl-3,5-dideoxy-D-glycero-\alpha-D-galacto-2-nonulopyranosylonate]-(2→3)-O-(2,4,6-tri-O-benzoyl-β-D-galactopyranosyl)-(1→3) - O-(2-acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl) - $(1 \rightarrow 4)$ -O-[methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero - α -D-galacto -2-nonulopyranosylonate) - (2 \rightarrow 3)] - O-(2,6-di - Oacetyl-β-D-galactopyranosyl)-(1→4)-2,3,6-tri-O-acetyl-β-D-glucopyranoside (7). Removal of benzyl groups and subsequent O-acetylation of 5 (100 mg, 0.037 mmol), as described for 6, gave 7 (70 mg, 76%) as an amorphous mass: $[\alpha]p$ -13.3° (c 1.4, CHCl₃); IR (KBr) 3300 (NH), 1750 and 1230 (ester), 1650 and 1540 (amide), 860 and 840 (Me₃Si), and 710 cm⁻¹ (Ph); ¹H NMR (CDCl₃) δ 0.89 (m, 2H, Me₃SiCH₂CH₂), 1.85-2.19 (21s, 63H, 17AcO, 4AcN), 2.52 (m, 1H, H-3feq), 2.84 (m, 1H, H-3geq), 3.45 and 3.80 (2s, 6H, 2MeO), 5.50 (d, 1H, $J_{3,4} = 3.4$ Hz, H-4c), 5.71 $(d, 1H, J_{3,4} = 2.9 Hz, H-4d), 6.03 (d, 1H, J_{NH,5} = 6.4 Hz, NH), 7.15-8.12 (m, 15H,$ 3Ph).

Anal. Calcd for C₁₂₁H₁₅₆N₄O₆4Si (2718.6): C, 53.46; H, 5.78; N, 2.06. Found: C, 53.22; H, 5.77; N, 2.05.

O-[Methyl 5- Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy - D - glycero - α - D - galacto-2 - nonulopyranosylono - 1',9 - lactone) - 4,7-di-O-acetyl-3,5-dideoxy -D -glycero-α -D -galacto-2 - nonulopyranosylonate] - (2→3) -O - (2,4,6-tri - O - benzoyl - β -D-galactopyranosyl) - (1→3)-O-(2-acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl) - (1→4) -O-(2,3,6-tri-O-acetyl-β-D-galactopyranosyl) - (1→4) -2,3,6-tri-O-acetyl-D-glucopyranose (8). A solution of 6 (110 mg, 0.05 mmol) in CH₂Cl₂ (0.5 mL) and trifluoroacetic acid (1 mL) was stirred for 30 min at room temperature. Ethyl acetate (1 mL) was added to the mixture and concentrated. Column chromatography (25:1 CH₂Cl₂-MeOH) of the residue on silica gel (10 g) gave 8 (77 mg, 75%) as an amorphous mass: IR (KBr) 3400-3100 (OH and NH), 1740 and 1230 (ester), 1650 and 1540 (amide), and 710 cm⁻¹ (phenyl).

Anal. Calcd for C98H₁₁9N₃O₅₃ (2187.0): C, 53.82; H, 5.48; N, 1.92. Found: C, 53.54; H, 5.29; N, 1.82.

O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylono-1',9-lactone) - 4,7-di -O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate]-(2→3)-O-(2,4,6-tri-O-benzoyl-β-D-galactopyranosyl)-(1→3)-O-(2-acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl)-(1→4)-O-[methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)-(2→3)]-O-(2,6-di-O-acetyl-β-D-galactopyranosyl)-(1→4)-2,3,6-tri-O-acetyl-D-glucopyranose (9). Selective removal of the 2-(trimethylsilyl)ethyl group of 7 (70 mg, 0.028 mmol) with trifluoroacetic acid (1 mL) in CH₂Cl₂ (2 mL) according to the method described for 8 gave 9 (56 mg, 83%) as an amorphous mass: IR (KBr) 3400-3200 (OH and NH), 1750 and 1230 (ester), 1650 and 1540 (amide), and 710 cm⁻¹ (Ph).

Anal. Calcd for C₁₁₆H₁₄₄N₄O₆₄ (2618.4): C, 53.21; H, 5.54; N, 2.14. Found: C, 53.05; H, 5.36; N, 1.97.

O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylono-1',9-lactone)-4,7-di-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate]- $(2\rightarrow3)$ -O-(2,4,6-tri-O-benzoyl-β-D-galactopyranosyl)- $(1\rightarrow3)$ -O-(2-acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl)- $(1\rightarrow4)$ -O-(2,3,6-tri-O-acetyl-β-D-galactopyranosyl)- $(1\rightarrow4)$ -2,3,6-tri-O-acetyl-α-D-glucopyranosyl Trichloroacetimidate (10). To a solution of 8 (250 mg, 0.11 mmol) in CH₂Cl₂ (2 mL) and trichloroacetonitrile (0.4 mL) was added DBU (19 μL, 0.13 mmol) at 0 °C, and the mixture was stirred for 1 h at 0 °C, then concentrated. Column chromatography (30:1 CH₂Cl₂-MeOH) of the residue on silica gel (20 g) gave 10 (240 mg, 96%) as an amorphous mass: [α]_D +18.5° (c 1.0, CHCl₃); IR (KBr) 3200 (NH), 1740 and 1230 (ester), 1650 and 1540 (amide), and 710 cm⁻¹ (phenyl); ¹H NMR MeO), 6.43 (d, 1H, J_{1,2} = 3.7 Hz, H-1a), 7.18-8.10 (m, 15H, 3Ph), 8.68 (s, 1H, C=NH).

Anal. Calcd for C₁₀₀H₁₁₉Cl₃N₄O₅₃ (2331.4): C, 51.52; H, 5.15; N, 2.40. Found: C, 51.46; H, 5.07; N, 2.26.

O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3.5dideoxy - D - glycero - a - D - galacto - 2 - nonulopyranosylono - 1',9 - lactone) -4.7-di-O-acetyl-3.5-dideoxy-D-glycero- \alpha - D-galacto-2-nonulopyranosylonate]- $(2\rightarrow 3)$ -O-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)- $(1\rightarrow 3)$ -O-(2acetamido - 4,6 - di - O- acetyl - 2 - deoxy - β - D - galactopyranosyl) - $(1 \rightarrow 4)$ - O-[methyl 5-acetamido-4.7.8.9 - tetra-O-acetyl-3.5 - dideoxy-D-glycero-a-Dgalacto-2-nonulopyranosylonate)- $(2 \rightarrow 3)$] -O-(2,6-di-O-acetyl- β -D-galactopyranosyl) - (1 → 4) -2,3,6 -tri-O-acetyl-α-D-glucopyranosyl Trichloroacetimidate (11). Treatment of 9 (56 mg, 0.023 mmol) with trichloroacetonitrile (0.1 mL) and DBU (4 µL) in CH₂Cl₂ (1 mL), then work up as described for 10 gave 11 (53 mg, 89%) as an amorphous mass: $[\alpha]_D + 5.7^\circ$ (c 1.1, CHCl₃); IR (KBr) 3300 (NH), 1750 and 1230 (ester), 1650 and 1540 (amide), and 710 cm⁻¹ (phenyl); ¹H NMR (CDCl₃) δ 1.85-2.19 (21s, 63H, 17AcO, 4AcN), 2.52 (m, 1H, H-3feq), 2.86 (m, 1H, H-3geq), 3.47 and 3.80 (2s, 6H, 2MeO), 5.71 (d, 1H, $J_{3.4} = 3.0$ Hz, H-4d), 6.06 (d, 1H, $J_{NH.5} = 6.4$ Hz, NH), 6.48 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1a), 7.27-8.12 (m, 15H, 3Ph) and 8.66 (s, 1H, C=NH).

Anal. Calcd for C₁₁₈H₁₄₄Cl₃N₅O₆₄ (2762.78): C, 51.30; H, 5.25; N, 2.53. Found: C, 51.29; H, 5.11; N, 2.31.

O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy - D-glycero - α - D-galacto-2 - nonulopyranosylono - 1',9 - lactone) - 4,7-di-O-acetyl-3,5-dideoxy-D-glycero - α-D-galacto-2-nonulopyranosylonate] - (2→3) - O-(2,4,6-tri-O-benzoyl-β-D-galactopyranosyl) - (1→3) - O-(2-acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl) - (1→4) - O-(2,3,6-tri-O-acetyl-β-D-galactopyranosyl) - (1→4) - O-(2,3,6-tri-O-acetyl-β-D-glucopyranosyl) - (1→1) - (2S,3R,4E) - 2-azido-3-O-benzoyl-4-octadecene-1,3-diol (14). To a solution of 10 (240 mg, 0.1 mmol) and (2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecene-1,3-diol (29 (12; 87 mg, 0.2 mmol) in CH₂Cl₂ (1.2 mL) were added molecular sieves 4Å (AW-300; 0.85 g), and the mixture was stirred for 5 h at room temperature, then cooled to 0 °C. Trimethylsilyl trifluoromethanesulfonate (TMSOTf; 40 μL, 0.2 mmol) was added, and the mixture was stirred for 10 h at 0 °C, then filtered. The insoluble materials were washed with M NaHCO₃ and H₂O, dried (Na₂SO₄) and concentrated. Column chromatography (30:1 CH₂Cl₂-MeOH) of the residue on silica gel

(30 g) gave **14** (150 mg, 53%) as an amorphous mass: $[\alpha]_D$ -0.6° (c 1.0, CHCl₃); IR (KBr) 3200 (NH), 3000-2900 (Me, methylene), 2200 (azide), 1730 and 1230 (ester), 1650 and 1540 (amide), and 710 cm⁻¹ (phenyl); ¹H NMR (CDCl₃) δ 0.88 (t, 3H, JMe,CH₂ = 7.0 Hz, MeCH₂), 1.24 (s, 22H, 11CH₂), 1.65 (t, 1H, J_{gem} = 13.0 Hz, H-3fax), 1.88-2.17 (17s, 51H, 14AcO, 3AcN), 2.52 (m, 1H, H-3eeq), 3.27 (s, 3H, MeO), 5,34 (d, 1H, J_{7,8} = 8.6 Hz, H-7f), 5.70 (d, 1H, J_{3,4} = 3.3 Hz, H-4f), 5.86 (m, 1H, H-5 of sphingosine), 7.21-8.11 (m, 20H, 4Ph).

Anal. Calcd for C₁₂₃H₁₅₆N₆O₅₅ (2598.6): C, 56.85; H, 6.05; N, 3.23. Found: C, 56.81; H, 6.01; N, 3.03.

O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5dideoxy - D - glycero - a - D - galacto - 2 - nonulopyranosylono - 1',9 - lactone) -4,7-di-O-acetyl-3,5-dideoxy-D-glycero-a-D-galacto-2-nonulopyranosylonate] - $(2 \rightarrow 3)$ - O-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl) - $(1 \rightarrow 3)$ - O-(2acetamido-4,6 - di - O - acetyl - 2 - deoxy - β - D-galactopyranosyl) - $(1 \rightarrow 4)$ - O-[methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-a-Dgalacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$]-O-(2,6-di-O-acetyl- β -D-galactopyranosyl) - $(1 \rightarrow 4)$ - O - (2,3,6 - tri - O - acetyl - β - D - glucopyranosyl) - $(1 \rightarrow 1)$ -(2S,3R,4E)-2-azido-3-O-(tert-butyldiphenylsilyl)-4-octadecene-1,3-diol (15). Glycosylation of (2S,3R,4E)-2-azido-3-O-(tert-butyldiphenylsilyl)-4-octadecene-1,3-diol³¹ (13; 23 mg, 40 µmol) with 11 (53 mg, 20 µmol) in CH₂Cl₂ (0.5 mL) in the presence of TMSOTf (15 µL) and MS-4Å (300 mg) for 10 h at 0 °C, then workup as described for 14 gave 15 (40 mg, 65%) as an amorphous mass: $[\alpha]_D$ -1.3° (c 0.8, CHCl₃); IR (KBr) 3200 (NH), 3100-2950 (Me, methylene), 2200 (azide), 1740 and 1230 (ester), 1650 and 1540 (amide), 710 and 700 cm⁻¹ (phenyl); ¹H NMR (CDCl₃) δ 0.88 (t, 3H, J_{Me,CH2} = 6.4 Hz, MeCH2), 1.05 (s, 9H, Me₃C), 1.27 (s, 22H, 11CH₂), 1.85-2.19 (21s, 63H, 17AcO, 4AcN), 2.52 (m, 1H, H-3feq), 2.85 (m, 1H, H-3geq), 3.47 and 3.80 (2s, 6H, 2MeO), 5.71 (d, 1H, $J_{3.4} = 3.5$ Hz, H-4d), 7.27-8.12 (m, 25H, 5Ph).

Anal. Calcd for C₁₅₀H₁₉₅N₇O₆₅Si (3164.3): C, 56.94; H, 6.21; N, 3.10. Found: C, 56.83; H, 6.08; N, 2.95.

O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy - D - glycero-α - D - galacto-2 - nonulopyranosylono - 1',9 - lactone) - 4,7-di-O-acetyl-3,5-dideoxy - D - glycero-α - D - galacto-2-nonulopyranosyl-

onate] - $(2 \rightarrow 3)$ - O - (2,4,6-tri-O-benzoyl - β - D - galactopyranosyl) - $(1 \rightarrow 3)$ - O- $(2-acetamido-4,6-di-O-acetyl-2-deoxy-\beta-D-galactopyranosyl)-(1 \rightarrow 4)-O (2,3,6-\text{tri}-O-\text{acetyl}-\beta-D-\text{galactopyranosyl})-(1\rightarrow 4)-O-(2,3,6-\text{tri}-O-\text{acetyl}-\beta-D$ glucopyranosyl) - $(1 \rightarrow 1)$ - (2S, 3R, 4E) - 3 - O - benzoyl - 2 - octadecanamido - 4 octadecene-1.3-diol (16). Hydrogen sulfide was bubbled through a stirred solution of 14 (150 mg, 0.054 mmol) in aq 83% pyridine (10 mL) for 3 days at 0 °C and concentrated. To a solution of the residue in CH₂Cl₂ (2.5 mL) were added octadecanoic acid (33 mg, 0.16 mmol) and 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (WSC: 27 mg, 0.16 mmol) and the mixture was stirred for 12 h at room temperature. After completion of the reaction, CH2Cl2 (50 mL) was added and the solution was washed with water, dried (Na2SO4) and concentrated. chromatography (25:1 CH2Cl2-MeOH) of the residue on silica gel gave 16 (84 mg, 52%) as an amorphous mass: [a]D +8.4° (c 1.0, CHCl3); IR (KBr) 3200 (NH), 3000-2900 (Me, methylene), 1740 and 1240 (ester), 1650 and 1540 (amide), and 710 cm⁻¹ (phenyl); ¹H NMR (CDCl₃) δ 0.90 (t, 6H, J_{Me,CH_2} = 6.8 Hz, 2MeCH₂), 1.24 (s, 52H, 26CH₂), 1.87-2.18 (17s, 51H, 14AcO, 3AcN), 2.50 (m, 1H, H-3eeg), 3.26 (s, 3H, MeO), 5.80 (m, 1H, H-5 of sphingosine), 7.20-8.10 (m, 20H, 4Ph).

Anal. Calcd for C₁₄₁H₁₉₂N₄O₅₆ (2839.1): C, 59.65; H, 6.82; N, 1.97. Found: C, 59.50; H, 6.68; N, 1.70.

O-[Methyl 5-Acetamido-8-O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylono-1',9-lactone)-4,7-di-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate]- $(2\rightarrow 3)$ -O-(2,4,6-tri-O-benzoyl-β-D-galactopyranosyl)- $(1\rightarrow 3)$ -O-(2-acetamido-4,6-di-O-acetyl-2-deoxy-β-D-galactopyranosyl)- $(1\rightarrow 4)$ -O-[methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 3)$]-O-(2,6-di-O-acetyl-β-D-galactopyranosyl)- $(1\rightarrow 4)$ -O-(2,3,6-tri-O-acetyl-β-D-glucopyranosyl)- $(1\rightarrow 1)$ -(2S,3R,4E)-3-O-(tert-butyldiphenylsilyl)-2-octadecanamido-4-octadecene-1,3-diol (17). To a solution of 15 (40 mg, 14 μmol) in benzene (1 mL) and water (0.04 mL) was added triphenylphosphine (7 mg, 28 μmol), and the mixture was stirred for 24 h at 30 °C and concentrated. To a solution of the residue in CH₂Cl₂ (0.8 mL) were added octadecanoic acid (11 mg, 42 μmol) and WSC (9 mg, 42 μmol), and the mixture was stirred for 15 h at

room temperature. The mixture was diluted with CH₂Cl₂ (50 mL) and the solution was washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (20:1 CH₂Cl₂-MeOH) of the residue on silica gel (10 g) gave **17** (41 mg, 95%) as an amorphous mass: $[\alpha]_D$ -6.3° (c 0.8, CHCl₃); IR (KBr) 3200 (NH), 3100-2950 (Me, methylene), 1740 and 1240 (ester), 1650 and 1540 (amide), 710 and 700 cm⁻¹ (phenyl); 1H NMR (CDCl₃) δ 0.88 (t, 6H, J_{Me},CH₂ = 6.4 Hz, 2*Me*CH₂), 1.05 (s, 9H, Me₃C), 1.26 (s, 52H, 26CH₂), 1.86-2.19 (21s, 63H, 17AcO, 4AcN), 2.51 (m, 1H, H-3eeq), 2.83 (m, 1H, H-3geq), 3.47 and 3.82 (2s, 6H, 2MeO), 5.73 (d, 1H, H-4d), 7.29-8.14 (m, 25H, 5Ph).

Anal. Calcd for C₁₆₈H₂₃₁N₅O₆₆Si (3404.8): C, 59.27; H, 6.84; N, 2.06. Found: C, 59.03; H, 6.58; N, 1.86.

Ganglioside GD1c (18). To a solution of 16 (84 mg, 28 μ mol) in MeOH (5 mL) was added NaOMe (10 mg), and the mixture was stirred for 24 h at 40 °C. Water (0.5 mL) was added and the solution was stirred for 10 h at 40 °C, neutralized with Amberlite IR-120 (H⁺) resin and filtered. The resin was washed with MeOH and the combined filtrate and washings were concentrated. Column chromatography (5:5:1 CHCl3-MeOH-H2O) on Sephadex LH-20 gave 18 (50 mg, quantitative) as an amorphous mass: [α]D +5.4° (c 1.0, 5:5:1 CHCl3-MeOH-H2O); ¹H NMR ((CD3)2SO-D2O) δ 0.88 (t, 6H, JMe,CH₂ = 6.4 Hz, 2MeCH₂), 1.24 (s, 52H, 26CH₂), 1.78-1.98 (3s, 9H, 3AcN), 2.42 and 2.87 (2m, 2H, H-3e,feq), 5.48 (m, 1H, H-4 of sphingosine), 5.60 (m, 1H, H-5 of sphingosine).

Anal. Calcd for C₈₄H₁₄₈N₄O₃₉ (1838.1): C, 54.89; H, 8.12; N, 3.05. Found: C, 54.60; H, 8.05; N, 3.04.

Ganglioside GT1a (19). To a solution of 17 (41 mg, 13 μmol) in MeCN (1.5 mL) was added M tetrabutylammonium fluoride in tetrahydrofuran (0.3 mL), and the mixture was stirred for 24 h at room temperature, then concentrated. To a solution of the residue in MeOH (2 mL) was added NaOMe (10 mg) and the mixture was stirred for 48 h at 40 °C. Potassium hydroxide (0.2 M, 0.2 mL) was added and the solution was stirred for 10 h at 40 °C, neutralized with Amberlite IR-120 (H⁺) resin and filtered. The resin was washed with MeOH, and combined filtrate and washings were concentrated. Column chromatography (5:5:1 CHCl3-MeOH-H₂O) of the residue on Sephadex LH-20 gave 19 (20 mg, 75%) as an amorphous mass: [α]_D +2.0° (c 0.6, 5:5:1 CHCl3-MeOH-H₂O); ¹H

NMR ((CD₃)₂SO-D₂O) δ 0.88 (t, 6H, J_{Me,CH₂} = 6.3 Hz, 2MeCH₂), 1.24 (s, 52H, 26CH₂), 1.76-1.88 (4s, 12H, 4AcN), 2.37-2.84 (m, 3H, H-3-e-geq), 4.16 (d, 1H, J_{1,2} = 9.6 Hz, H-1a), 4.28 (m, 2H, H-1b,d), 4.85 (d, 1H, J_{1,2} = 7.3 Hz, H-1c), 5.32 (m, 1H, H-4 of sphingosine), 5.58 (m, 1H, H-5 of sphingosine).

Anal. Calcd for C95H₁₆₅N₅O₄₇ (2129.4): C, 53.59; H, 7.81; N, 3.29. Found: C, 53.36; H, 7.79; N, 3.11.

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REFERENCES

- 1. Presented at the XVIIIth International Carbohydrate Symposium, Milan, Italy, July 21-26, 1996.
- (a) R. Kuhn and H. Wiegandt, Z. Naturforsch., 19b, 256 (1964); (b) E. Klenk and M. Naoi, Hoppe Seylers Z. Physiol. Chem., 349, 288 (1968).
- 3. (a) R. Kuhn, H. Wiegandt and H. Egge, Angew. Chem., 73, 580 (1961); (b) R. Kuhn and H. Wiegandt, Chem. Ber., 96, 866 (1963).
- 4. I. Ishizuka and H. Wiegandt, Biochim. Biophys. Acta, 260, 279 (1972).
- (a) S. Ando, Y. Hirabayashi, K. Kon, F. Inagaki, S. Tate and V.P. Whittaker, J. Biochem. (Tokyo), 111, 287 (1992); (b) Y. Hirabayashi, T. Nakao, F. Irie, V.P. Whittaker, K. Kon and S. Ando, J. Biol. Chem., 267, 12973 (1992).
- (a) S. Tsuji, T. Yamakawa, M. Tanaka and Y. Nagai, J. Neurochem., 50, 414 (1988);
 (b) S. Tsuji, S. Yamashita and Y. Nagai, J. Biochem. (Tokyo), 104, 498 (1988).
- 7. Y. Hirabayashi, A. Hyogo, T. Nakao, K. Tsuchiya, Y. Suzuki, M. Matsumoto, K. Kon and S. Ando, J. Biol. Chem., 265, 8144 (1990).
- (a) M. Tiemeyer, Y. Yasuda and R.L. Schnaar, J. Biol. Chem., 264, 1671 (1989);
 (b) M. Tiemeyer, P. S.-Hill and R.L. Schnaar, J. Biol. Chem., 265, 6239 (1990).
- 9. C.M. Gammon, K.K. Vaswani and R.W. Ledeen, Biochemistry, 26, 6239 (1987).
- T. Hama, Y. Kushima, M. Miyamoto, M. Kubota, N. Takei and H. Hatanaka, Neuroscience, 40, 445 (1991).
- 11. R.L. Hopfer, S.W. Johnson, M. Masserini, A. Giuliani and J.A. Alhadeff, *Biochem. J.*, 266, 491 (1990).

- 12. S. Tsuji, M. Arita and Y. Nagai, J. Biochem. (Tokyo), 94, 303 (1983).
- 13. A. Kameyama, H. Ishida, M. Kiso and A Hasegawa, Carbohydr. Res., 200, 269 (1990).
- 14. A. Hasegawa, T. Nagahama, H. Ohki and M. Kiso, J. Carbohydr. Chem., 11, 699 (1992)
- 15. H. Prabahanjan, K. Aoyama, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, 233, 87 (1992).
- 16. A. Hasegawa, H.-K. Ishida, T. Nagahama and M. Kiso, J. Carbohydr. Chem., 12, 703 (1993).
- 17. K. Hotta, H. Ishida, M. Kiso and A. Hasegawa, J. Carbohydr. Chem., 13, 665 (1994).
- 18. A. Kameyama, H. Ishida, M. Kiso and A. Hasegawa, J. Carbohydr. Chem., 13, 641 (1994).
- 19. T. Terada, M. Kiso and A. Hasegawa, Carbohydr. Res., 259, 201 (1994).
- 20. H.-K. Ishida, Y. Ohta, Y. Tsukada, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, **246**, 75 (1993).
- 21. H.-K. Ishida, Y. Ohta, Y. Tsukada, Y. Isogai, H. Ishida, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, 252, 283 (1994).
- 22. H.-K. Ishida, H. Ishida, M. Kiso and A. Hasegawa, Carbohydr. Res., 260, c1 (1994).
- 23. H.-K. Ishida, H. Ishida, M. Kiso and A. Hasegawa, *Tetrahedron: Asymmetry*, 5, 2493 (1994).
- 24. Z. Bartoszewicz, J. Koscielak and T. Pacuszka, Carbohydr. Res., 151, 77 (1986).
- 25. S. Ando and R. K. Yu, J. Biol. Chem., 252, 6247 (1977).
- 26. A. Hasegawa, K. Hotta, A. Kameyama, H. Ishida and M. Kiso, J. Carbohydr. Chem., 10, 439 (1991).
- K. Jansson, S. Ahlfors, T. Frejd, J. Kihlberg, G. Magnusson, J. Dahmén, G. Noori and K. Stenvall, J. Org. Chem., 53, 5629 (1988).
- 28. R.R. Schmidt and G. Grundler, Synthesis, 885 (1981).
- (a) R.R. Schmidt and P. Zimmermann, Angew. Chem. Int. Ed. Engl., 25, 725 (1986); (b) M. Kiso, A. Nakamura, Y. Tomita and A. Hasegawa, Carbohydr. Res., 158, 101 (1986); (c) Y. Ito, M. Kiso and A. Hasegawa, J. Carbohydr. Chem., 8, 285 (1989).
- 30. T. Adachi, Y. Yamada, I. Inoue and M. Saneyoshi, Synthesis, 45 (1977).
- 31. T. Ehara, A. Kameyama, Y. Yamada, H. Ishida, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, 281, 237 (1996).
- 32. M. Mori, Y. Ito and T. Ogawa, Carbohydr. Res., 195, 199 (1990).
- 33. T. Fujisawa, T. Mori, K. Fukumoto and T. Sato, Chem. Lett., 1891 (1982).